



THOUSAND WORDS ABOUT...

DOI: <https://doi.org/10.20883/jms.2017.263>

The molecular basis of non-syndromic orofacial clefts and tooth agenesis

Agnieszka Danuta Gaczowska, Paweł Piotr Jagodziński, Adrianna Mostowska

Department of Biochemistry and Molecular Biology, Poznan University of Medical Sciences, Poznan, Poland

ABSTRACT

Non-syndromic orofacial clefts and tooth agenesis are two of the most common craniofacial birth defects. Both of them have a complex etiology, with genetic and environmental factors involved. Additionally, the epigenetic modifications have been implicated in the pathogenesis of these structural malformations. Despite an increasing number of research studies, using a variety of methodological approaches, the role of genetic factors in the etiology of orofacial clefts and tooth agenesis is still not well elucidated. The most consistent findings across studies concerning the genetic factors influencing the risk to orofacial clefts include the association of polymorphic variants of the *IRF6* gene and the chromosomal locus 8q24.21. The major candidate gene for tooth agenesis in the European populations is *WNT10A*; its pathogenic mutations are present in more than 50% of patients with this dental anomaly. It has been found that both orofacial clefts and tooth agenesis, which co-occurrence is often reported, may share common candidate genes.

Keywords: orofacial clefts, tooth agenesis, etiology, candidate genes.

Introduction

Non-syndromic orofacial clefts (OFC) and tooth agenesis (TA) are two of the most common craniofacial birth defects [1, 2]. OFC affect 1 per 700 live births in the global population [1]. According to the *Polish Registry of Congenital Malformations, the prevalence of orofacial clefts in Poland ranges from 1/500 to 1/1000 births* [www.rejestrwad.pl]. OFC are divided into two main forms: non-syndromic cleft lip with or without cleft palate and cleft palate only [3]. The incidence of TA, excluding the lack of the third molars, varies from 1.6 to 9.6% depending on ethnic background [2]. TA can be classified based on the number of missing teeth into hypodontia (the lack of one to five teeth), oligodontia (the lack of 6 or more teeth) and anodontia (the complete absence of teeth). In this classification, the third molars are not taken into account since their absence is highly prevalent [2]. The co-occurrence of OFC and TA is often reported [4, 5]. Patients with OFC have an increased risk of dental anomalies, including

alteration in tooth number, size, shape, a timing of formation and eruption comparing to the general population [6]. It has been shown that dental anomalies appear primarily in the cleft area and their prevalence is higher in left-sided OFC [7]. Additionally, in patients with OFC the agenesis of teeth *outside the cleft area* have also been reported to be more frequent [8]. This observation may indicate that the same molecular mechanisms may be shared in the development of the teeth, palate, and lip [8].

The etiology of non-syndromic OFC and TA is complex with genetic and environmental components [3, 9]. Additionally, the epigenetic modifications have been implicated in the pathogenesis of these structural malformations [10]. Genetic studies using a variety of research approaches, including linkage studies, candidate gene analyses, and genome-wide association studies, have identified a number of genes and chromosomal regions underlying these craniofacial anomalies [9, 11]. However, nucleotide variants of identified

candidate genes and chromosomal loci can still explain only a fraction of the predicted heritability. It has been demonstrated that both OFC and TA have a number of common candidate genes, which nucleotide variants can influence their risk [4, 5].

Across OFC' studies conducted in various populations, including the Polish population, the most consistent results were observed for nucleotide variants located in the *IRF6* gene (OMIM *607199) and the chromosomal region 8q24.21 [12–14]. The *IRF6* gene encodes a transcription factor, which is involved in the regulation of the keratinocyte proliferation-differentiation switch and formation of oral periderm [15]. It is worth noting that in a study conducted in the Latvian population, the *IRF6* variant (rs642961) located in the promoter region was found to be more frequent in individuals presenting OFC associated with tooth agenesis when compared to healthy individuals [16]. Moreover, Vieira *et al.* have demonstrated that this functional variant, disrupting an AP-2 α binding site in the *IRF6* enhancer, is associated with the risk of isolated TA [12, 17, 18]. The 8q24.21 risk locus, identified by the first genome-wide association study conducted for OFC and further confirmed by a number of post-GWAS replication studies, is a gene-poor region devoid of protein-coding genes [19, 20]. Studies using mice as model organisms have demonstrated that this chromosomal locus contains very distant *cis*-acting enhancers controlling the expression of the *Myc* gene during craniofacial development [21]. Mice homozygous for the deletion including this medianasal enhancer region show mild alterations in the face morphology and *occasionally* cleft lip and palate [21]. Within the 8q24.21 chromosomal region, which nowadays is considered as a key susceptibility locus for non-syndromic OFC, the top marker associated with the risk of this anomaly is rs987525 [19]. A significant association between this intragenic variant and the co-occurrence of OFC and TA outside the cleft region was also observed [22].

The major candidate genes for non-syndromic TA include *WNT10A* (OMIM *606268), *MSX1* (OMIM *142983), *PAX9* (OMIM *167416), *AXIN2* (OMIM *604025), *EDA* (OMIM *300451) and *EDAR* (OMIM *604095). Van den Boogaard *et al.* [23] have demonstrated that the *WNT10A* mutations are present in more than 50% isolated TA cases. Pathogenic mutations within the coding region of *WNT10A* have also been identified in 62% of tooth agenesis patients from the Polish population [24]. The *WNT10A* gene is a member of the Wnt family, which consists of genes encoding secreted signaling proteins involved in a number of developmental

processes during embryogenesis [25]. Interestingly, the missense mutation of the *WNT10A* gene has been associated with the increased risk of non-syndromic OFC in the Chinese population [26]. Moreover, nucleotide variants in *WNT3* (OMIM *165330), *WNT3A* (OMIM *606359), *WNT5A* (OMIM *164975), *WNT9A* (OMIM *602863), and *WNT11* (OMIM *603699) have been found to be significantly associated with non-syndromic orofacial clefts in various populations [27, 28]. Similarly, polymorphisms and mutations in the *MSX1* gene are known factors increasing the risk of non-syndromic OFC [29]. In addition, *MSX1* and two other major TA candidate genes, *PAX9* and *AXIN2*, have been associated with the co-occurrence of cleft anomalies and TA [5]. The *MSX1* and *PAX9* genes encode transcription factors that play an essential role during embryogenesis [9]. It has been demonstrated that these genes are co-expressed during craniofacial development, and the genetic interactions between their protein products are involved in the regulation of the lip formation and tooth morphogenesis [30]. *Msx1* and *Pax9* deficient mice lack all teeth, which development is arrested at the bud stage, and exhibit a number of craniofacial defects, including cleft palate [31, 32]. The *AXIN2* gene encodes a protein which is a negative regulator of the Wnt-signalling pathway [33].

Besides the genes described above, there are a number of other candidate genes and chromosomal loci underlying the co-occurrence of OFC and TA. The systemic review conducted by Phan *et al.* [5] revealed that they include among others the TGF pathway genes and the cancer predisposing gene *CDH1* (OMIM *192090).

In summary, OFC and TA are one of the most common craniofacial anomalies that share a number of common candidate genes. There is growing evidence suggesting that tooth agenesis should be considered as an extended phenotype for oral clefts [34].

Acknowledgements

Conflict of interest statement

The authors declare no conflict of interest.

Funding sources

The study was supported by the Polish National Science Centre, grant no. 2012/07/B/NZ2/00115.

References

1. Mossey PA, Little J, Munger RG, Dixon MJ, Shaw WC. Cleft lip and palate. *Lancet*. 2009 Nov 21;374(9703):1773–1785.

2. Polder BJ, Van't Hof MA, Van der Linden FP, Kuijpers-Jagtman AM. A meta-analysis of the prevalence of dental agenesis of permanent teeth. *Community Dent Oral Epidemiol.* 2004 Jun;32(3):217–226.
3. Dixon MJ, Marazita ML, Beaty TH, Murray JC. Cleft lip and palate: understanding genetic and environmental influences. *Nat Rev Genet.* 2011 Mar;12(3):167–178.
4. Vieira AR. Oral clefts and syndromic forms of tooth agenesis as models for genetics of isolated tooth agenesis. *J Dent Res.* 2003 Mar;82(3):162–165.
5. Phan M, Conte F, Khandelwal KD, Ockeloen CW, Bartzela T, Kleefstra T, et al. Tooth agenesis and orofacial clefting: genetic brothers in arms? *Hum Genet.* 2016 Dec;135(12):1299–1327.
6. Tannure PN, Oliveira CA, Maia LC, Vieira AR, Granjeiro JM, Costa Mde C. Prevalence of dental anomalies in nonsyndromic individuals with cleft lip and palate: a systematic review and meta-analysis. *Cleft Palate Craniofac J.* 2012 Mar;49(2):194–200.
7. Bartzela TN, Carels CE, Bronkhorst EM, Kuijpers-Jagtman AM. Tooth agenesis patterns in unilateral cleft lip and palate in humans. *Arch Oral Biol.* 2013 Jun;58(6):596–602.
8. Slayton RL, Williams L, Murray JC, Wheeler JJ, Lidral AC, Nishimura CJ. Genetic association studies of cleft lip and/or palate with hypodontia outside the cleft region. *Cleft Palate Craniofac J.* 2003 May;40(3):274–279.
9. Yin W, Bian Z. The Gene Network Underlying Hypodontia. *J Dent Res.* 2015 Jul;94(7):878–885.
10. Wang J, Sun K, Shen Y, Xu Y, Xie J, Huang R, et al. DNA methylation is critical for tooth agenesis: implications for sporadic non-syndromic anodontia and hypodontia. *Sci Rep.* 2016 Jan 13;6:19162.
11. Leslie EJ, Marazita ML. Genetics of cleft lip and cleft palate. *Am J Med Genet C Semin Med Genet.* 2013 Nov;163C(4):246–258.
12. Rahimov F, Marazita ML, Visel A, Cooper ME, Hitchler MJ, Rubini M, et al. Disruption of an AP-2alpha binding site in an IRF6 enhancer is associated with cleft lip. *Nat Genet.* 2008 Nov;40(11):1341–1347.
13. Mostowska A, Hozyasz KK, Wojcicki P, Biedziak B, Paradowska P, Jagodzinski PP. Association between genetic variants of reported candidate genes or regions and risk of cleft lip with or without cleft palate in the polish population. *Birth Defects Res A Clin Mol Teratol.* 2010 Jul;88(7):538–545.
14. Thieme F, Ludwig KU. The Role of Noncoding Genetic Variation in Isolated Orofacial Clefts. *J Dent Res.* 2017 Oct;96(11):1238–1247.
15. Richardson RJ, Dixon J, Jiang R, Dixon MJ. Integration of IRF6 and Jagged2 signalling is essential for controlling palatal adhesion and fusion competence. *Hum Mol Genet.* 2009 Jul 15;18(14):2632–2642.
16. Krasone K, Lāce B, Akota I, Care R, Deeley K, Kūchler EC, et al. IRF6 AP-2a binding site promoter polymorphism is associated with oral clefts in Latvia. *Stomatologija.* 2014;16(4):132–136.
17. Vieira AR, Modesto A, Meira R, Barbosa AR, Lidral AC, Murray JC. Interferon regulatory factor 6 (IRF6) and fibroblast growth factor receptor 1 (FGFR1) contribute to human tooth agenesis. *Am J Med Genet A.* 2007 Mar 15;143A(6):538–545.
18. Vieira AR, McHenry TG, Daack-Hirsch S, Murray JC, Marazita ML. Candidate gene/loci studies in cleft lip/palate and dental anomalies finds novel susceptibility genes for clefts. *Genet Med.* 2008 Sep;10(9):668–674.
19. Birnbaum S, Ludwig KU, Reutter H, Herms S, Steffens M, Rubini M, et al. Key susceptibility locus for nonsyndromic cleft lip with or without cleft palate on chromosome 8q24. *Nat Genet.* 2009 Apr;41(4):473–477.
20. Murray T, Taub MA, Ruczinski I, Scott AF, Hetmanski JB, Schwender H, et al. Examining markers in 8q24 to explain differences in evidence for association with cleft lip with/without cleft palate between Asians and Europeans. *Genet Epidemiol.* 2012 May;36(4):392–399.
21. Uslu VV, Petretich M, Ruf S, Langenfeld K, Fonseca NA, Marioni JC, et al. Long-range enhancers regulating Myc expression are required for normal facial morphogenesis. *Nat Genet.* 2014 Jul;46(7):753–758.
22. Yildirim M, Seymen F, Deeley K, Cooper ME, Vieira AR. Defining predictors of cleft lip and palate risk. *J Dent Res.* 2012 Jun;91(6):556–561.
23. van den Boogaard MJ, Créton M, Bronkhorst Y, van der Hout A, Hennekam E, Lindhout D, et al. Mutations in WNT10A are present in more than half of isolated hypodontia cases. *J Med Genet.* 2012 May;49(5):327–331.
24. Mostowska A, Biedziak B, Zadurska M, Dunin-Wilczynska I, Lianeri M, Jagodzinski PP. Nucleotide variants of genes encoding components of the Wnt signalling pathway and the risk of non-syndromic tooth agenesis. *Clin Genet.* 2013 Nov;84(5):429–440.
25. MacDonald BT, Tamai K, He X. Wnt/beta-catenin signaling: components, mechanisms, and diseases. *Dev Cell.* 2009 Jul;17(1):9–26.
26. Feng C, Duan W, Zhang D, Zhang E, Xu Z, Lu L. C392T polymorphism of the Wnt10a gene in non-syndromic oral cleft in a northeastern Chinese population. *Br J Oral Maxillofac Surg.* 2014 Oct;52(8):751–755.
27. Chiquet BT, Blanton SH, Burt A, Ma D, Stal S, Mulliken JB, et al. Variation in WNT genes is associated with non-syndromic cleft lip with or without cleft palate. *Hum Mol Genet.* 2008 Jul 15;17(14):2212–2218.
28. Mostowska A, Hozyasz KK, Biedziak B, Wojcicki P, Lianeri M, Jagodzinski PP. Genotype and haplotype analysis of WNT genes in non-syndromic cleft lip with or without cleft palate. *Eur J Oral Sci.* 2012 Feb;120(1):1–8.
29. Jezewski PA, Vieira AR, Nishimura C, Ludwig B, Johnson M, O'Brien SE, et al. Complete sequencing shows a role for MSX1 in non-syndromic cleft lip and palate. *J Med Genet.* 2003 Jun;40(6):399–407.
30. Nakatomi M, Wang XP, Key D, Lund JJ, Turbe-Doan A, Kist R, et al. Genetic interactions between Pax9 and Msx1 regulate lip development and several stages of tooth morphogenesis. *Dev Biol.* 2010 Apr 15;340(2):438–449.
31. Satokata I, Maas R. Msx1 deficient mice exhibit cleft palate and abnormalities of craniofacial and tooth development. *Nat Genet.* 1994 Apr;6(4):348–356.
32. Peters H, Neubüser A, Kratochwil K, Balling R. Pax9-deficient mice lack pharyngeal pouch derivatives and teeth and exhibit craniofacial and limb abnormalities. *Genes Dev.* 1998 Sep 1;12(17):2735–2747.
33. Jho EH, Zhang T, Domon C, Joo CK, Freund JN, Costantini F. Wnt/beta-catenin/Tcf signaling induces the tran-

scription of Axin2, a negative regulator of the signaling pathway. *Mol Cell Biol.* 2002 Feb;22(4):1172–1183.

34. Letra A, Menezes R, Granjeiro JM, Vieira AR. Defining subphenotypes for oral clefts based on dental development. *J Dent Res.* 2007 Oct;86(10):986–991.

Acceptance for editing: 2017-11-10
Acceptance for publication: 2017-12-23

Correspondence address:

Adrianna Mostowska, PhD
Department of Biochemistry and Molecular Biology
Poznan University of Medical Sciences
6 Swieczickiego Street, 60-781 Poznan, Poland
phone: +48 618546511
fax: +48 618546510
email: amostowska@wp.pl